Journal Of Harmonized Research (JOHR)

Journal Of Harmonized Research in Applied Sciences 6(1), 2018, 01-08



ISSN 2321 - 7456

Original Research Article

DEVELOPMENT AND VALIDATION OF HPLC METHOD FOR THE IGURATIMOD

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Abstract: Analytical method with *accuracy*, precision and stability indicting was developed for Iguratimod, achieved the required chromatographic separation by High pressure liquid chromatography (HPLC) technique.

HPLC method with accuracy, precision, robustness and stability indicating was developed and optimized for quantitative evaluation of Iguratimod as active pharmaceutical ingredient. Chromatographic separation for Iguratimod and its related compounds was achieved by using analytical column with stationary phase Inertsil ODS-3 (Dimension: length 150mm, internal diameter 4.6mm and particle size 5μ) and mobile phase (Buffer and acetonitrile in gradient proportion) at flow rate 1.0ml/minute with detection at 257nm. Squared correlation coefficient (r²) was observed more than 0.999 with concentration window of 12.5 to 75μ g/ml. Iguratimod was retained at about 19.0±2.0 minutes. Forced degradation study for Iguratimod was carried out under the stress condition to get required degradation by using acid, base, thermal, photolytic and oxidation by hydrogen peroxide. Degradation products which are generated during stress condition did not interfere to active ingredient,

Iguratimod. This analytical method optimized, validated for all the parameters specified in ICH guideline, to get results selective, accurate and precise in presence of degradation impurities.

Keywords: Iguratimod, validation, development, stability indicating methodology.

Introduction: Chemically, Iguratimod 3-Formylamino-7-methylsulfonylamino-6-henoxy-4H-

For Correspondence: ravindra.nehete@ipca.com. Received on: January 2018 Accepted after revision: January 2018 Downloaded from: www.johronline.com DOI: https://doi.org/10.30876/johr.6.1.2018.1-8 1-benzopyran-4-one is used as an anti-inflammatory drug for the treatment of rheumatoid arthritis. It has following structure,



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IUPAC name for Iguratimod is N-[(formylamino)-4-oxo-6-phenoxy-4Hchromen-

7-yl] methane sulfoanamide. Iguratimod was first reported in product patent US4954518.[1] Its Therapeutic category is Anti-arthritic and novel immunomodulator.[2] Iguratimod is a nuclear factor NF-kB activation inhibitor used in the treatment of rheumatoid arthritis. It also suppressed inflammatory cytokine production in cultured human synovial cells induced by tumor necrosis factor (TNF)- α by inhibiting the activity of nuclear factor-kB. Several synthesis processes are reported for Iguratimod. [3-6]. Efficacy of a drug substance is critical for its safety assessment. It is compulsory to identify and characterize the possible impurities in active drug. This compound is aromatic heterocyclic compound; belong to class of organic compound known as chromones. These are compounds containing a benzopyran-4-one moiety.

Separation of active drug and possible impurities was determined by high performance liquid chromatography instrumental technique. Since literature did not cite such methodology for determination of Iguratimod as well as its degradants, it was planned to develop a user friendly, selective, accurate and precise HPLC based methodology for determination of Iguratimod in presence of possible impurities.

Chromatographic technique is used in most of the cases for analysis of active drugs as this technique is specific, accurate, precise, and user friendly. Chromatographic technique gets rid of tedious extraction and isolation procedures.

separation Chromatographic is multistage separation technique, in which the sample components are distributed in two phase's i.e. stationary phase and mobile phase. Stationary phase is either solid or a liquid supported on a solid or a gel. Stationary phase packed in column, spread as layer and distributed as a thin film, or applied by appropriate technique. Mobile phase may be either gas or liquid / Separation is based on supercritical fluid. adsorption, mass distribution (partition), or ion exchange; due differences or to in physicochemical properties of the molecules, like molecule size, mass of molecule, and volume of molecules. This type of chromatography is used for qualitative and quantitative analysis by using different techniques.

Method and Requirements:

Instrument: HPLC, make: Waters, pump, UV detector, column oven, injector, etc.

Chromatographic column: Inertsil ODS-3 (Dimension: 150 x 4.6mm, 5µ)

Chemicals and reagents: Potassium dihydrogen orthophosphate (Analytical grade) Acetonitrile (HPLC grade)

Orthophosphoric acid (Analytical grade) Water (HPLC grade)

Mobile phase preparation: Mobile phase-A: 20mM potassium dihydrogen orthophosphate with pH 3.50 ± 0.05 with 10% orthophosphoric acid, filter this mobile phase by using 0.45μ membrane filter and Sonicate to degas.

Mobile phase-B: Acetonitrile

Diluent preparation: In equal proportion, mix Mobile phase-A and Mobile phase-B.

Standard stock solution: Weigh accurately about 50 mg of standard of Iguratimod and transfer into 50 ml volumetric flask. Dissolve in about 30ml diluent and makeup the volume up to the mark with diluent (1000ppm solution).

Working standard solution: The standard stock solution was used to prepare working standard solutions of concentrations 12.5, 25, 37.5, 50, 62.5 and 75 ppm. Solution having drug concentration of 50 ppm was used as a working standard for stress degradation studies. Standard and sample solution with concentration of 50ppm, were analyzed at 257nm and recorded the chromatograms (Fig. 2).

Sample solution: Weigh accurately about 50 mg of sample of Iguratimod to be tested and transfer into 50 ml volumetric flask. Dissolve in about 30ml diluent and makeup the volume up to the mark with diluent (1000ppm solution).

Further dilutions were made to get the final stock having concentration equivalent to 50ppm.

Various trials were made for optimization of chromatographic conditions to finalize the chromatographic parameters and conditions like mobile phase, its ratio and flow rate. Finally, the one giving the best results were optimized. The chromatographic estimation of Iguratimod and its separation from degradation products was achieved using analytical column Inertsil ODS-3 (Dimension: Length 150 mm, 4.6 mm internal diameter and particle size 5μ) with mobile phase-A (Phosphate buffer pH 3.50 ± 0.05) and Acetonitrile, flow rate of 1.0 ml/min. The UV detection was done at 257 nm.

Validation of proposed method: Validated the proposed methodology as per ICH guidelines, for the parameters like specificity, linearity, precision, accuracy and robustness etc.[11, 12].

System suitability testing: Five replicates of drug concentration of 50ppm were injected and recorded the chromatograms to check with the system suitability parameters [9].

Forced degradation studies: Forced degradation study was performed for Iguratimod as per ICH Q1A (R2) for Acid and base, oxidation and thermal stress conditions. photostability study as per ICH Q1B. The stress conditions employed were 1M Hydrochloric acid for acid hydrolysis, 0.1 M Sodium Hydroxide for base hydrolysis, 10% Hydrogen Peroxide for oxidative hydrolysis. Iguratimod samples were subjected for thermal degradation and photolytic degradation [9,10].

Acid hydrolysis: degradation by acid was performed by adding 1 M Hydrochloric acid in stock solution (1000ppm) of Iguratimod. This solution was subjected to stress condition of 60°C for 0.5 hours. The resulting solutions were neutralized with 1 M Sodium Hydroxide and further diluted to obtain the concentration of 50µg/ml.

Base hydrolysis: Base induced forced degradation was performed by adding an aliquot of stock solution (1000ppm) of Iguratimod to 0.1M Sodium Hydroxide. The resulting solutions was neutralized with 0.1 Μ Hydrochloric acid and further diluted to obtain the concentration of 50µg/ml.

Oxidation stress: To study the effects of oxidative conditions, aliquot of stock solution (1000ppm) of Iguratimod was added to 10% Hydrogen Peroxide solution with stressed condition as heating at 60°C for 4 hours, further diluted this solution to get 50ppm solution.

Thermal stress: Test solid was heated to 105°C for 24 hours to study effect of temperature. Solution was prepared to get final concentration as 50ppm.

Photolytic stress: Standard stock solution (1000ppm) of Iguratimod was subjected to UV exposure in UV chamber for about 24 hours, further diluted to obtain final concentration of 50ppm.

Results and Discussion:

Selection of solvent: Since Iguratimod was soluble in mixture of water and acetonitrile, it was used as a diluent. For dilution purpose single solvents like water or acetonitrile was not used as it does not complete solubility and results in hazy solution.

Optimization of mobile phase: Final mobile phase Buffer and acetonitrile was selected since trials done using methanol: Water caused peak merging with other impurities. Gradient ratio was optimized because altering the ratio to isocratic mode, resulted in peak merging and peak shape deterioration. The flow rate 1.0 ml/min was optimized since at lower flow rates, i.e., at 0.8 ml/min or 1.2 ml/min, peak merging was observed.

 Table 1: Mobile phase preparation and gradient

program

program							
Mobile phase preparation							
Mobile	20mM potassium dihydrogen						
phase-A	orthophosphate, adjust pH to 3.50 \pm						
:	0.05 with 10% Orthophosphoric						
acid. Filter and degas							
Mobile							
phase-B	Acetonitrile (HPLC grade)						
:							

Gradient program :						
Time	Mobile phase-A	Mobile phase-B				
0	80	20				
4	68	32				
22	68	32				
27	20	80				
32	20	80				
35	80	20				
45	80	20				

Optimization of chromatographic conditions Purpose of analytical method optimization is specifically to identify the analyte peak in presence possible impurities. The chromatographic separation of Iguratimod from its degradants was achieved using Inertsil ODS-3 (Dimension: Length 15 cm, 4.6 mm internal diameter and particle size 5μ) with mobile phase in gradient proportion at flow rate of 1.0 ml/min and detection wavelength of 257 nm.

Table 2: Optimized chromatographic parameters

and conditions					
	Chromatographic				
Parameters	conditions				
	Inertsil ODS-3, 150mm				
Stationary phase	x 4.6mm, 5µ				
Flow rate (Gradient)	1.0ml/min				
Injection volume	10µ1				
Detection wavelength	UV 257 nm				
Runtime	45.0 minutes				
Column oven					
temperature	25°C				
	Mobile phase-A :				
Diluent	Mobile phase-B (50:50)				

System suitability testing: The result for system suitability is shown in below Table 3. Result found within the acceptable limit. Hence the system was suitable for the proposed method.



Figure 2: HPLC chromatogram obtained during simultaneous determination of system suitability.

Chromatographic conditions: using Inertsil ODS-3 (Dimension: Length 150 mm, 4.6 mm internal diameter and particle size 5μ ; flow rate 1.0 mL /min; mobile phase acetonitrile and phosphate buffer (gradient proportion) and UV detection at 257 nm.

Table 3: System suitability data

System suitability (% Relative standard						
	dev	iation)				
Paramet RSD for Observed result (n=5) Accept ance criteria						
Repeata bility (%RSD)	0.19	% RSD <2.0	Method passes system suitability criteria			

Validation

Linearity: The linearity was observed in concentration range from 12.5ppm to 75ppm for Iguratimod. The regression line equation was plotted between concentration and peak area. The regression equation y = 59739.8082x - 9969.3093 was obtained from the linearity data. The squared correlation coefficient was found to be 0.9995.



Figure 3: Linearity plot Table 4: Linearity data

Linearity for Iguratimod				
Conc.(ppm)	Area response			
12.50	743131			
25.06	1480182			
37.49	2224179			
50.12	2960983			

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62.65	3790740
75.18	4451972
Slope =	59739.8082
Intercept =	-9969.3093
Squared correlation coefficient =	0.9995
Correlation coefficient =	0.9998

Specificity: The results of specificity are shown in Figure 4. No interference was observed due to blank and degradants present in Iguratimod.

Precision: Analytical results were summarized in Table 9. The % RSD was found within the acceptable limit, i.e., <2.

Table-5: System precision

System Precision										
No. Area			Average		Standard deviation		standard	deviation		
Injection	-1	29499	958							
Injection	-2	29624	85	2959839		5	5520 F		0.	
Injection	-3	29618	867			2 8	556.5 703	1	1	
Injection	-4	29628	394			8		9)	
Injection	-5	29619	989							
Tab	le-6:	Metho	d pro	ecisio	n (An	al	yst-1)			
	Met	thod Pr	ecisi	on: A	Analy	st-	1			
Test		Area	Average	area	% Assay		Average assay (%)	SD	%RSD	
Sample -1	296 295	55910 59665	296	52788	99.2					
Sample -2	294 294	48426 48928	294	8677	99.4					
Sample -3	296 297	54305 77197	297	0751	100.	3	9.	585	38	
Sample -4	294 294	41362 47634	294	4498	99.4		66	0.30	0.(
Sample -5	295 294	52583 48867	295	0725	99.5					
Sample -6	296 296	65660 62466	296	64063	99.7					

Table-7: Method precision (Analyst-2)

						,	
Method Precision : Analyst-2							
Test	Area	Aver age area	% A	ssay	assay	SD	%RSD
Sample-1	2644169	2645	99	99.7			
	2040914	242	+				
Sample-2	2634320	2033 433	99	9.4			
	2650622	2649					
Sample-3	2648907	765	99	9.9	L.	33	8
Commute 4	2620388	2635	00	5	99.	.18	0.1
Sample-4	2649945	167	99	9.5			
Sample 5	2647953	2648	00	9 8			
Sample-3	2649359	656	75	7.0			
Sample-6	2651313	2651	90	99.8			
Sample 0	2652084	699	//				
Table-8:	Intermediat	e prec	isior	n (Cur	nula	tive	e
Intones 1'-	re Descision	sults)					
Intermedia	te Precision						
lyst	st	say	age)	%		S	ative)
Anal	Te	% As	(Aver	'erall	say	D	8SD umula
			-	Ŏ.	AS		(Cl %I
	Sample-1	99.	2	1			
	Sample-2	99.	4]			
	Sample-3	100).3				
lyst	Sample-4	99.	4	1			
Anal	Sample-5	99.	5	99.6		0	
	Sample-6	99.	7			2	0.28
	Sample-1	99.	7			28	0.20
	Sample-2	99.	4			5	
t-2	Sample-3	99.	9	-			
lys	Sample-4	99.	5				
Ana	Sample-5	99.	8	-			
\prec	Sample 6	00	Q D	1			

Recovery studies: The % recovery was observed within the acceptable limits, i.e., 100.0%, 99.5% and 100.1% at the levels of 50%, 100% and 150% respectively. The results are summarized in Table 9.

99.8

Sample-6

Robustness: Robustness of the method was studied by deliberate changes in chromatographic condition and parameters like flow rate, column oven temperature and pH of mobile phase. No significant impact observed on results due to change in flow rate, column oven temperature and pH of mobile phase.

Forced degradation studies: Iguratimod was subjected to various stress conditions like acid / base hydrolysis, oxidation, photolytic and thermal stress conditions as per ICH guidelines. The results are tabulated in Table 10.



Figure 4: As such test (Without treatment) and diluent blank

Acid and base hydrolysis: Adequate degradation was achieved under acid and base degradation stress condition. Degradation in the applied stress conditions of the acid and base hydrolysis are 7.8% and 4.6% respectively for Iguratimod. The chromatograms showed the presence of degraded products (Figures 5 and 6).



Figure 5: Acid degradation and diluent blank



Figure 6: Base degradation and diluent blank **Oxidation degradation:** Iguratimod was found to be quite stable in oxidation stress condition. It showed 1.5% for Iguratimod (Figure 7).



Figure 7: Oxidation degradation and diluent blank

Thermal degradation: Iguratimod was found to be quite stable in thermal stress condition, i.e. 0.4% for Iguratimod (Figure 8).



Photolytic degradation: Iguratimod was found to be quite stable in photolytic stress condition, and degraded sufficiently up to 1.3% with applied stress condition (Figure 9).



Table 9:	Validation	summary
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Validation summary							
Parameters		Results					
Range (50% to 150%)	12.50 ppm to 75.0ppm						
Regression line equation	59739.808	2 x – 996	9.3093				
Slope	59739.808	2					
Intercept	9969.3093						
Squared							
correlation	0.9995						
coefficient							
Correlation	0.0000						
coefficient	0.9998						
04 A courses	%Recove	0/	DSD				
%Accuracy	ry	70	KSD				
50	100.0	0.14					
100	99.5	0.08					
150	100.1	0.09					
Precision	Concentr						
TICCISION	ation	/0	KSD				
Repeatability	50 ppm						
n=5	50 ppm	0.19					
Intraday	50 ppm						
precision n=6	50 ppm	0.08					
Interday	50 ppm						
precision n=6	50 ppm	0.18					
Cumulative	50 nnm						
results $(n = 12)$	50 ppm	0.28					
Robustness : N	lo significar	nt change	s observed				
with deliberate c	hanges in m	ethod par	ameters				
Table 10: Stress	condition a	nd degrad	lation data				
		%Degrad					
Stress condition			ation				
Initial (As such)			NA				
0.1M, 5ml S	droxide						
solution, 0.0 hrs.	4.6						
1M, 5ml Hydro							
at 60°C, 0.5 hrs	7.8						
10.0% 5ml Hydrogen peroxide, heat							
60°C for 4.0 hrs	1.5						
24.0 hrs test solu	1.3						
Test Solid hea	Test Solid heated at 105°C for						
24.0hrs	0.4						

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Conclusions: The present studies are very much useful for prediction of stability behavior of Iguratimod as per the ICH guidelines. Iguratimod was found to be more stable under stress conditions. The method was found to be accurate and precise with good and consistent recoveries at all levels studied. This indicates there is no interference of degradants as well as other impurities for determination of drug content by this methodology.

RSD was also less than 2% showing high degree of precision of the proposed method. This method of analysis is accurate, precise, rapid and cost-effective. The proposed method can be used for routine analysis for Iguratimod as drug substances and can be a very good too for quality control in bulk manufacturing.

References

1. Shuntaro Takano, Chosaku Yoshida, Takihiro Inaba, Keiichi Tanaka, Ryuko Takeno, Hideyoshi Nagaki, Tomoya Shimotori, 4H-1benzopyran-4-one derivative or its salt, process for producing the same and pharmaceutical composition comprising the same as active ingredient., *Toyama Chemical Company*, Ltd., Tokyo, Japan US4954518, 4 Sept, 1990.

2. Wang Jinyi , Li Xudong , Lin Guoqiang , Zhang Zheng Gen , Wang Lin , Lu Wen bud Preparation of 3-(formamide)-7-(methylsulfonyl amine)-6-(phenoxy)-4H-1-(benzopyran)-4-

ketone., Jiangsu Yangtze River Pharmaceutical Group Co. Ltd., CN 1462748.aceutical Group Co. Ltd., CN 1462748.

3. Takihiro Inaba, keiichi Tanaka, ruuko takeno, hideyoshi nagaki, Chosaku Yoshida, Shuntaro takano, Synthesis and Antiinflammatory Activity of 7-Methanesulfonylamino-6phenoxychromones. Antiarthritic Effect of the 3-Formylamino Compound (T-614) in Chronic inflammatory disease models.*Chem. Pharma. Bull*, 2000; 48(1): 131-139.

4. Shanghai Huagong, 2008; 32(12): 22-24.

5. Wang Yan Xiang, Gao Hong, Cao Feng hua, Song Dan Qing, Synthesis of Iguratimod *Zhongguo Xinyao Zazhi*, 2006; 15(23): 2042-2044.

6. Huagong_Shikan, 2010; 24 (9): 267[1]).

7. ICH, Q1A (1993) Stability testing of new drug substances and products in Proceedings of the international conference on harmonization. Geneva, Switzerland.

8. ICH, Q2B (1996) Harmonised tripartite guideline, Validation of analytical procedure Methodology, International conference on harmonization. Geneva, Switzerland.

9. ICH (2002) Guidance on analytical method validation, International convention on quality for the pharmaceutical industry. Toronto, Canada.

10. ICH, Q1B (1996) Stability testing: photostability testing of new drug substances and Products in International Conference on Harmonization. IFPMA, Geneva, Switzerland.

11. ICH (1996) Validation of analytical procedures methodology ICH harmonized tripartite guidelines.

12.General Chapter, Validation of compendial methods, United States Pharmacopeia, 26th Revision, National Formulary, 21st Edition, Rockville, MD, The United States Pharmacopoeial Convention, Inc, 2440; 2003.

13. International Conference on Harmonization (ICH) of Technical Requirements for the Registration of Pharmaceuticals for Human Use, Validation of analytical procedures, ICH-Q2A, Geneva; 1995.

14. International Conference on Harmonization (ICH) of Technical Requirements for the Registration of Pharmaceuticals for Human Use, Validation of analytical procedures: Methodology, ICH-Q2B, Geneva; 1996.

15. US FDA Technical Review Guide: Validation of Chromatographic Methods, Center for Drug Evaluation and Research (CDER), Rockville, MD; 1993.

16. US FDA, General principles of validation, Rockville, MD, Center for Drug Evaluation and Research (CDER); 1987.

17. US FDA, Guidelines for submitting samples and analytical data for method validation, Rockville, MD, Center for Drugs and Biologics Department of Health and Human Services; 1987.